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ACE-2 Expression in the Small Airway Epithelia of Smokers and COPD Patients:

Implications for COVID-19

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The World Health Organization (WHO) has declared coronavirus disease 2019 (COVID-19) as a pandemic [1]. COVID-19 is caused by severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2). COVID-19 displays symptoms ranging from mild to severe (pneumonia) that can lead to death in some individuals [2-4]. As of March 24, 2020, there have been 422,566 cases of COVID-19 worldwide and 18,887 deaths [5]. SARS-CoV-2 uses the angiotensin converting enzyme II (ACE-2) as the cellular entry receptor [6]. While the virus can infect individuals of any age, to date, most of the severe cases have been described in those over the age of 55 years and with significant comorbidities such as chronic obstructive pulmonary disease (COPD) [7]. Here, we determined whether patients with COPD have increased expression of ACE-2 in bronchial epithelial cells in lower respiratory tract.

Patients undergoing bronchoscopy at St. Paul's Hospital (SPH), Vancouver, Canada for clinical purposes were enrolled. The protocol was approved by the University of British Columbia/Providence Health Care Ethics Board (UBC/PHC REB H15-02166). All patients were required to be 19 years of age or older, who underwent spirometry according to international guidelines[8]. Patients with COPD were defined as those having a clinical diagnosis of COPD made by a board-certified respiratory physician and either a forced expiratory volume in 1 second (FEV1)/forced vital capacity (FVC) <70% or clear evidence of emphysema on computed tomographic (CT) imaging on visual inspection. Cytologic brushings were obtained in subsegmental airways (6th-8th generation) of the lung that were unaffected by the patient's underlying clinical indication for bronchoscopy.

Total RNA was extracted from cytologic brushings using the RNeasy Mini Kit (Qiagen, Hilden, Germany). Transcriptomic sequencing was performed on the NovaSeq 6000 (Illumina, San Diego, CA) at a sequencing depth of 55 million reads. Raw sequencing reads were quality controlled with FastQC[9] and aligned to the GENCODE (version 31) GRCh37 genome reference using STAR (Spliced Transcripts Alignment to a Reference) [10]. After alignment, the data were quantified using RSEM (RNA-Seq by Expectation Maximization) to obtain the read counts. Limma voom[11] was applied to normalize the counts to log2 counts per million reads (CPM), which was used in the downstream analysis.

Two cohorts were used for validation; the details of which are provided in a previous publication [12]. First, we used 16 datasets obtained from bronchial brushings of 10th-12th generation bronchi collected at a single center; transcriptome measurement was performed using the U133 Plus 2.0 microarray (denoted as the Cornell Dataset)[13]. Second, we used dataset GSE37147 consisting of bronchial brushings from the 6th-8th generation airways with gene

expression profiles generated from the GeneChip Human Gene 1.0 ST microarray[14]. This dataset was denoted as British Columbia Cancer Agency (BCCA) cohort.

We also determined protein expression of ACE-2 in resected lung tissue specimens. These samples were obtained from 10 current smokers with COPD (FEV1/FVC 61±7%), 9 non-smoker controls (FEV1/FVC 85±2%), and 8 healthy current smokers (FEV1/FVC 78±6%). Human lung tissue samples were obtained with informed consent from patients undergoing thoracic surgery as part of the James Hogg Lung Registry (UBC/PHC REB Protocol Hoo-50110). Formalin-fixed paraffinembedded human lung tissues were stained with antibody against ACE-2 (Ab15348; Abcam) using the Bond Polymer Refine Red Detection kit on a Leica Bond Autostainer as previously described[15]. Airway epithelial-specific ACE-2 protein intensity was quantified using the Aperio imaging system with normalization to the length of the basement membrane (Leica Biosystem; Concord, Ontario).

For the primary study population, log2 CPM of ACE-2 was the principal outcome of interest. Robust linear models were used to determine whether 1) ACE-2 was differentially expressed in patients with COPD and in smokers after adjustment for age and sex and 2) ACE-2 expression was significantly correlated with lung function. All analyses were performed in R (version 3.5.0). In the immunohistochemistry dataset, Kruskal-Wallis with Dunn's Multiple Comparisons tests was used. Continuous data are expressed as mean±SD, unless otherwise indicated.

The average age of the SPH cohort was 64.8±12.0 years; 55% were females and 24% were current smokers. Compared to control subjects (N=21), those with COPD (N=21) had lower FEV1% (72.0±15.6 vs 85.9±17.9% predicted; p=0.011) and FEV1/FVC (64.1±7.9 vs 76.3±5.9%; 2.621×10⁻⁶). Most (79%) underwent bronchoscopy for investigation of lung nodules, followed by chronic cough (7%) and lymphadenopathy (7%). ACE-2 expression in the epithelial cells was significantly increased in COPD versus non-COPD subjects (COPD=2.52±0.66 versus non-COPD=1.70±0.51; p=7.62×10⁻⁴; Figure 1A). There was a significant inverse relationship between ACE-2 gene expression and FEV1% of predicted (r=-0.24; p=0.035; Figure 1B). Interestingly, smoking status was also significantly related to ACE-2 gene expression levels in airways of these participants with current smokers having a significantly higher gene expression than never smokers (current smokers=2.77±0.91 versus never smokers=1.78±0.39; p=0.024). Former smokers had gene expression levels inbetween that of never and current smokers (former smokers=2.00±1.23; Figure 1C). Conditional on the smoking status, the association between ACE-2 expression and COPD was still significant (Adjusted Mean±SE of non-COPD: 0.90±0.65 versus COPD: 1.75±0.82, p=0.016).

Next, we validated the above findings in: 1) the Cornell Cohort (N= 211) and 2) the BCCA cohort (N=238). The average age of the Cornell Cohort was 43.6±10.5 years with 33.2% of the cohort being females. There were 32.2% who were never smokers and 67.8% who were current smokers at the time of the bronchoscopy. The average age of the BCCA cohort was 64.5±5.9 years with 43.3% of the cohort being females. All were heavy smokers with at least 30 pack-years of smoking. Of these, 41.6% were current smokers at the time of the bronchoscopy and the remaining were former smokers.

In both the Cornell and BCCA cohorts, current smokers had increased ACE-2 gene expression levels in the airways compared with never smokers (in the Cornell cohort; current smokers= 4.34 ± 0.45 versus never smokers= 4.15 ± 0.36 ; p= 1.92×10^{-3}) and with former smokers (in the BCCA cohort; current smokers= 6.05 ± 0.53 versus former smokers= 5.57 ± 0.37 ; p< 2×10^{-16}). In the BCCA cohort, pre-bronchodilator FEV1 was measured and it was significantly related to ACE-2 gene expression level (r=-0.10; p=0.037).

Representative images of epithelial-specific ACE-2 protein expression in non-smokers, healthy smokers and smokers with COPD are shown in Figure 1D. ACE-2 expression in the human small airway epithelium was significantly increased in COPD compared to non-smokers but not in healthy smokers (Figure 1D). ACE-2 protein staining was largely restricted to the airway epithelium of COPD and cells in the submucosal compartment.

There is a worldwide outbreak of COVID-19 coronavirus. Although most patients infected and diagnosed with CVOID-19 disease have mild symptoms, approximately 20% of individuals have demonstrated severe or critically severe disease including symptoms and signs of pneumonia, respiratory failure, septic shock and multi-organ failure. The estimated case-fatality rate is 1-2% [2, 3]. Importantly, nearly all deaths have occurred in those with significant underlying chronic diseases including COPD and cardiovascular diseases [4]. The reason for this observation is largely unknown.

One possibility is differential expression of ACE-2, which is the main receptor used by SARS-CoV-2 to gain entry into the host mucosa and cause active infection. Here, we investigated gene expression levels of ACE-2 in the airways of individuals with and without COPD and found that COPD and current smokers had significantly increased expression of ACE-2. Importantly, gene expression levels of ACE-2 were inversely related to individual's FEV1, suggesting a dose-dependent response. These findings were observed in 3 different cohorts, indicating their generalizability and robustness.

ACE-2 is a type I transmembrane metallocarboxypeptidase with homology to angiotensin converting enzyme (ACE). In contrast to ACE, which converts angiotensin I to the active vasoconstrictor, angiotensin II, ACE-2 breaks down angiotensin II to its metabolites including angiotensin-(1–9) and angiotensin-(1–7), which are potent vasodilators, and thus may be a negative regulator of the renin-angiotensin system[16]. ACE-2 is expressed in a variety of different tissues including both the upper and lower respiratory tract, myocardium and the gastrointestinal mucosa [17]. Although its role in human health and disease has not been fully elucidated, it appears to have an important regulatory role in blood pressure and cardiac function. The physiologic role of ACE-2 in the airways is largely unknown. However, in mice, ACE-2 has been shown to protect animals from severe lung injury related to aspiration and sepsis [18].

To our knowledge, our study is the first to demonstrate increased ACE-2 expression in airways of current (but not former) smokers and those with COPD. These results are also consistent with previous observations in small animals wherein smoke exposure has been shown to upregulate both the expression and activity of ACE-2 in the airways [19, 20]. While the upregulation of ACE-2 may be useful in protecting the host against acute lung injury, chronically, this may predispose individuals to increased risk of coronavirus infections, which uses this receptor to gain entrance into epithelial cells. This may in part explain the increased risk of viral respiratory tract infection in active smokers and virus-related exacerbations in those with COPD.

There were limitations to the study. First, the study was cross-sectional and as such, we could not determine whether interventions such as inhaled corticosteroids or bronchodilators (for those with COPD) could modulate ACE-2 gene expression in the airways. Second, as receptor expression is one of many host factors that govern infection risk among individuals, the precise attributable risk (for coronavirus infections) imposed by cigarette smoking and COPD is uncertain. Third, although the airway epithelia is the major source of entry for COVID-19, the virus can gain host entry through other ports including gastrointestinal mucosa, which was not evaluated in this study. Fourth, we did not have access to upper airway tissues, which may also become infected with SARS-CoV-2.

In summary, active cigarette smoking and COPD up-regulate ACE-2 expression in lower airways, which in part may explain the increased risk of severe COVID-19 in these populations. These findings highlight the importance of smoking cessation for these individuals and increased surveillance of these risk subgroups for prevention and rapid diagnosis of this potentially deadly disease.

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Figure 1A: A violin plot of ACE-2 expression in small airways of COPD and non-COPD subjects in the St. Paul's Hospital Cohort.

The red box indicates the median and the interquartile range. The P-value was obtained from the robust linear model

Abbreviations: ACE-2, angiotensin converting enzyme II; COPD, chronic obstructive pulmonary disease; CPM, counts per million reads

Figure 1B: A scatter plot of ACE-2 expression in small airways according to FEV1 % Predicted in the St. Paul's Hospital Cohort.

Abbreviations: ACE-2, angiotensin converting enzyme II; COPD, chronic obstructive pulmonary disease; CPM, counts per million reads

ACE-2 gene expression in airway epithelia is inversely related to FEV1% predicted (p=0.0348)

Figure 1C. A violin plot of ACE2 expression in small airways of never, former and current smokers in the St. Paul's Hospital Cohort.

The red box indicates the median and the interquartile range. The P-value was obtained from the robust linear model.

Abbreviations: ACE-2, angiotensin converting enzyme II; CPM, counts per million reads

Figure 1D. Protein staining of ACE2 in airways of Individuals with and without COPD

Human kidney slide was the positive control for ACE-2. The specificity of the antibody against ACE-2 was determined using an immunoblot assay with HEK2 cell lysates as a positive control. The expected molecular weight of ACE-2 is 90-100 kDa.

In airways, most of the protein expression was noted in the epithelial layer, most pronounced in those with COPD.

Abbreviations: ACE-2, angiotensin converting enzyme II; NHBE, normal human bronchial epithelial cells; NS, non-smoker

